# **VOCATIONAL CURRICULUM - 2020**

# **Medical Lab Technician**



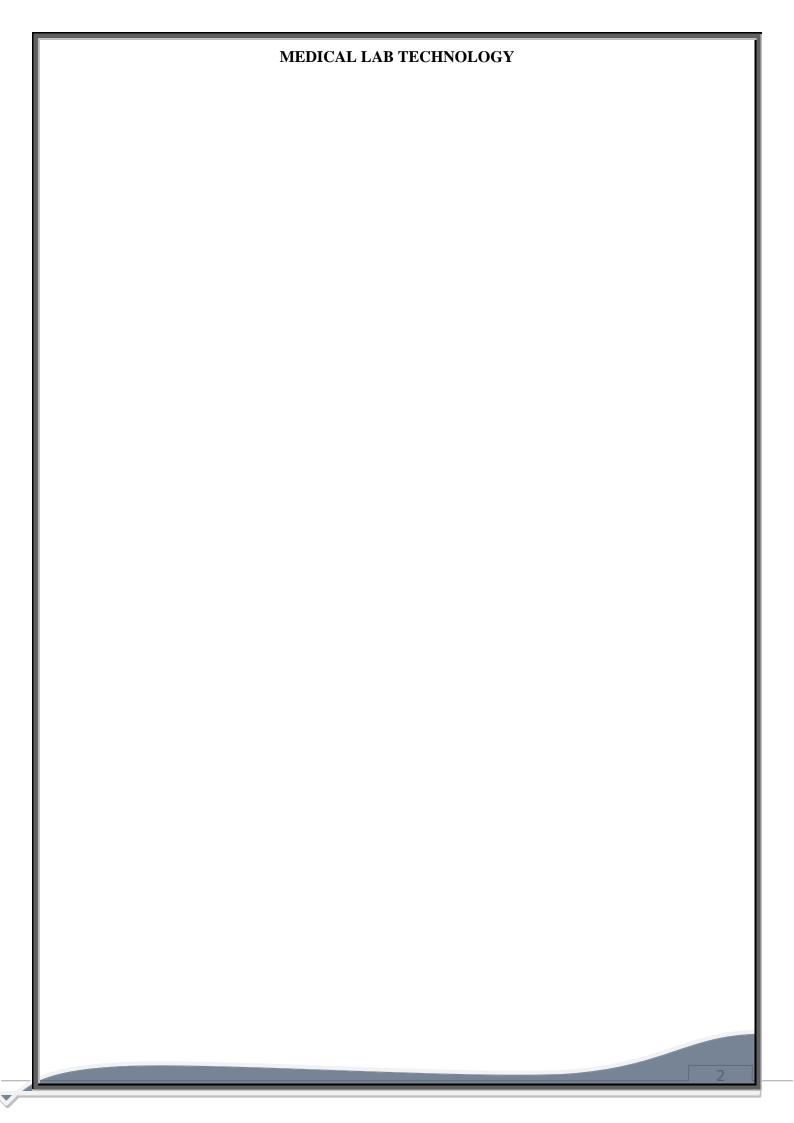
# **State Institute of Vocational Education**

O/o the Commissioner of Intermediate Education, Telangana State, Hyderabad



# **Board of Intermediate Education**

Telangana State, Hyderabad



Dr. A.Ashok I.A.S COMMISSIONER 500001



INTERMEDIATE EDCATION
Government of Telangana
Nampally, Hyderabad-

Phone: 040-24655915

#### Fore word

In any developing society with a booming population, Vocational Education occupies an important position for generating large scale employment opportunities. Viewed in this context the importance of Vocational Education for our country cannot be over emphasized. Vocationalization of Secondary Education was introduced in 1988 at the Intermediate level. Recently, the Government of India has developed a National Skills Qualification Framework for establishing a clear path for vocational education from the school level to the highest level. The Department of Intermediate Education has recently framed a new curriculum to bring greater value to the system of vocational education. The primary aim of this reform is to prepare the students with employable skills for absorption in organized sectors and in few cases, even for self-employment.

State Institute of vocational education and Board of Intermediate Education, Telangana have reviewed the curriculum of vocational courses in order to reorient them for their practical approach. Greater emphasis is now being placed on Laboratory work and on the job training.

Simultaneously, The State Institute of Vocational Education and the department of Intermediate Education are presently making efforts to upgrade the quality of infrastructure in the colleges to meet the challenges of the changed curriculum. I am confident that the revised curriculum and the new text books would prove to be beneficial to the students in the vocational stream and help them train in need based productive courses leading to gainful employment.

Commissioner of Intermediate Education Government of Telangana

S No.	Contents	Page
I	Introduction	No. 5
II	Objectives of the Course	5
III	Skills to be provided	5
IV	Job Opportunities	8
- '	a) Wage Employment	
	b) Self-Employment	
V	Scheme of Instruction and Examination	9
	SYLLABUS	1 -
VI	First year Intermediate	12
	Theory	
	Paper I: Bio-Chemistry -I	
	Paper II : Microbiology & Pathology	
	Paper III: Anatomy & Physiology	
	Practicals	
	Paper I: Bio-Chemistry -I	
	Paper II: Microbiology & Pathology	
	Paper III: Anatomy & Physiology	
VIII	Second Year Intermediate	22
	Theory Paper I: Bio-chemistry-II	
	Paper II: Micro-biology	
	Paper III: Pathology	
	Tupor III. Tuttiology	
	Practicals	
	Paper I: Bio-chemistry-II	
	Paper II: Micro-biology	
	Paper III: Pathology	
IX	Model Question Papers	37
X	List of Equipment	49
	a) Collaborating Institutions for curriculum transaction	
VI	b) On – the – Job Training Sites	52
XI	Teaching staff and their Qualification	53
XII	Vertical Mobility	53
XIII	Reference Books	53
XIV	List of Participants	54

#### I Introduction

Diagnostics play an important and vital role in the field of medicine. Without the aid of proper diagnostics, accurate conclusions cannot be drawn and suitable medical or surgical treatment cannot be given. Preparation of reagents of the purest quality also needs special attention and care. Thus Medical Lab Technology Courses is gaining greater importance. Clinical advances & studies in the fields of medicine, surgery, Pharmaceutical industries and nutrition also require technicians.

The Medical Laboratory Technician course is designed to train man power to effectively carry out medical Laboratory Technical work in various departments of medical, dental, pharmacy colleges, peripheral laboratories, research and diagnostic centers and also to set up and run own clinical laboratories.

#### **II.** Objectives of the Course

- To train the students to work in diagnostic labs.
- To train the students to work in manufacturing units of diagnostic regents.
- To train the students to assist the qualified experts in these fields.
- To train the students to understand the organization of Hospitals, Research Laboratories etc.
- To train the students to attend to analytical work and Research & Development [R&D] work in drug labs and pharmaceutical industries.
- To Train the student to attend basic Emergency, diagnostic requirement to the patient in Day and night Duties.

### III.Skills to be provided.

#### GENERAL SKILLS FOR FIRST YEAR MLT STUDENTS

- Identification of patient.
- Giving instructions to patients for 24 hour urine sample collection.
- Giving instructions to patients for GTT.
- Giving instructions to patients for semen sample collection.
- Giving instructions to patients for sputum sample collection.
- Giving instructions to patients for urine sample collection for culture.
- Collecting proper history relevant to the test.
- Preparation of patient for Phlebotomy.
- Technique of phlebotomy.
- Collecting sample in appropriate container.(Sample Collection)
- Preparing the sample for processing.
- Preservation and storage of specimen.
- Accurately processing of specimens.
- Appropriately discarding the specimens.
- Reception and registration of the sample
- Entering the report in register and dispatch of report...

- Handling of the Apparatus.
- Accurately measuring, weighing etc.
- Maintaining the electronic balance.
- Cleaning and sterilization of the apparatus.
- Disposal of Lab waste.
- Know Limitation of the Tests.

#### SKILLS IN PATHOLOGY FOR FIRST YEAR MLT STUDENTS

- Preparing the anticoagulants
- Preparing the Leishman's stain.
- Preparing peripheral blood smear.
- Staining of peripheral blood smear.
- Preparing and staining thick smear.
- Estimation of haemoglobin by Sahli's method and by photoelectric method.
- Estimation of ESR
- Estimation of PCV by Micro and Macro Methods.
- Physical examination of Urine.
- Chemical Examination of urine for sugars-manual method/strip method.
- Chemical Examination of urine for Ketone bodies-manual method/strip method.
- Chemical Examination of urine for Protiens-manual method/strip method.
- Chemical Examination of urine for blood-manual method/strip method.
- Chemical Examination of urine for bile salts -manual method/strip method.
- Chemical Examination of urine for bile pigments -manual method/strip method.
- Preparing wet smear for urine microscopy.
- Preparing processing the blood sample for reticulocyte count by wet and dry method.
- Charging the improved Neubauer chamber
- WBC Count by manual method.
- Platelet Count by manual method.
- Processing of body fluids for cell count.
- Processing of semen for sperm count.
- Coagulation Tests- Bleeding time, Clotting time, Prothrombin time [PT]& APTT
- Absolute Eosinophil count- Materials required, diluting fluids, procedure,
- Sickle cell preparation.
- Screening of blood donor.

#### SKILLS IN PATHOLOGY FOR SECOND YEAR MLT STUDENTS

- Performing Coombs Test a) Direct b) indirect
- Staining of Cytological smears.
- Setting up of tray for bone marrow aspiration
- Preparing bone marrow smears methods- Imprints smears, crush smears.
- Staining of bone marrow smear.
- Buffy coat preparation.
- Techniques of grouping and cross matching

- Receiving and preserving histopathology specimens.
- Maintaining the registers-receiving register, grossing register, slide register, report issue register.
- Taking the consent for investigation.
- Counseling of patient before and after test.
- Maintaining haematologyanalyser.
- Processing the sample in haematologyanalyser.
- Fixation of Histopathology specimen.
- Preparing the specimen for grossing.
- Processing of histopathology specimen manual and automated method.
- Paraffin embedding and block making
- Trimming of blocks
- Tissue section cutting.
- H&E Staining of tissue sections
- H&E Staining cytology slides.
- Mounting of slides
- Maintaining microtome and tissue processor and tissue floatation bath.
- Sharpening of microtome knife.
- Staining of histopathology sections.
- Fixatives for cytology specimens.
- Processing fluid sample for cytology.
- Preparing tray for FNAC and guided aspiration.
- PAP stain preparation and staining of smears.
- Maintenance & Preservation of cytology slides, histopathology blocks & slides.

#### SKILLS IN BIO-CHEMISRY FOR FIRST YEAR MLT STUDENTS

- Carefull Study of the requisition for the tests.
- Should learn to talk to patient politely and prepare them for blood drawing.
- Ability to identify the patient sample, and labeling and entry into the register.
- Transport of Sample to the Appropriate table for processing
- Should be able to handle the instruments with adequate care.
- Should be able to assist the senior technician in the lab.
- Should be able to dispose the Bio medical waste.
- Dispatch of the reports.

#### SKILLS IN BIO-CHEMISRY FORSECOND YEAR MLT STUDENTS

- Should be able to do estimations individually.(With Manual Procedures, Semi Auto Analyzers and Fully auto analyzer)
- Should be able to handle Semiauto analyzers.
- Should be able to handle automated pipettes.

#### SKILLS IN MICROBIOLOGY FOR FIRST YEAR MLT STUDENTS

• Reception area and its quality control.

- Cleaning and maintenance of equipment, glassware.
- Preparation of stains, culture media and sugar Medias.
- Disposal of lab waste

#### SKILLS IN MICROBIOLOGY FOR SECOND YEAR MLT STUDENTS

- Preparation of direct smears and staining of smears.
- Techniques of inoculation on media and biochemical sugars for the isolation of bacteria.
- Stool concentration techniques and microscopy of stool.
- Serological tests.
- Antibiotic sensitivity tests.
- Maintenance of stock cultures.
- Mycology processing

# IV. Job Opportunities.

#### A. Wage Employment

- 1. Laboratory technicians in various departments in diagnostic centers.
- 2.Laboratory technicians in hospitals of various sectors.
- 3. Laboratory technicians in various departments in medical and pharmacy Colleges.
- 4. Laboratory Technicians in Clinical studies in various laboratories like
  - i) National Institute of Nutrition.
  - ii) Central Drug Research Laboratory
  - iii) Molecular Biology Labs.
- 5.Laboratory Technicians in Quality Control, R&D Sectors of Drug Manufacturing units.

#### **B.** Self Employment

- 1. Preparation and sale of readymade reagent kits and media.
- 2. Distribution of Lab Chemicals, Glassware, Lab Instruments and their spare parts etc.

# V. SCHEME OF INSTRUCTION AND EXAMINATION

# ANNUAL SCHEME OF INSTRUCTION AND EXAMINATION **MEDICAL LAB TECHNICIAN**

#### FIRST YEAR

Part-A		Th	eory	Practicals		Total	
		Periods	Marks	Periods	Marks	Periods	Marks
1.	English	150	50	-	-	150	50
2.	General	150	50	-	-	150	50
	Foundation						
	course						
	Part-B						
3.	Paper-1	135	50	135	50	270	100
	Bio-Chemistry -I						
4.	Paper-II	135	50	135	50	270	100
	Microbiology &						
	Pathology						
5.	Paper-III	135	50	135	50	270	100
	Anatomy &						
	Physiology						
	OJT	-	-	365	100	365	100
	TOTAL	705	250	770	250	1475	500

<sup>\*</sup>on the Job Training for 1st year from 1st November to 31st December

#### **SECOND YEAR**

			SECON	DILAK			
	Dowl A	Theory		Practicals		Total	
Part-A		Periods	Marks	Periods	Marks	Periods	Marks
1.	English	150	50	-	-	150	50
2.	General	150	50	-	-	150	50
	Foundation						
	course						
Par	t-B						
3.	Paper-1	110	50	115	50	225	100
	Bio-chemistry-II						
4.	Paper-II	110	50	115	50	225	100
	Micro-biology						
5.	Paper-III	110	50	115	50	225	100
	Pathology						
Par	t-C						
6.	OJT	0	0	450	100	450	100
	Total	630	250	795	250	1425	500
TO	TAL FIRST YEAD	R AND SI	ECOND YI	EAR MARI	KS 1000	•	•

<sup>\*</sup>OJT Programme for 2<sup>nd</sup> year students from 1<sup>st</sup>August to 30<sup>th</sup>October

#### **EVALUATION OF ON THE JOB TRAINING:**

The "On the Job Training" shall carry 100 marks for each year and pass marks is 50. During on the job training the candidate shall put in a minimum of 90 % of attendance.

The evaluation shall be done in the last week of January.

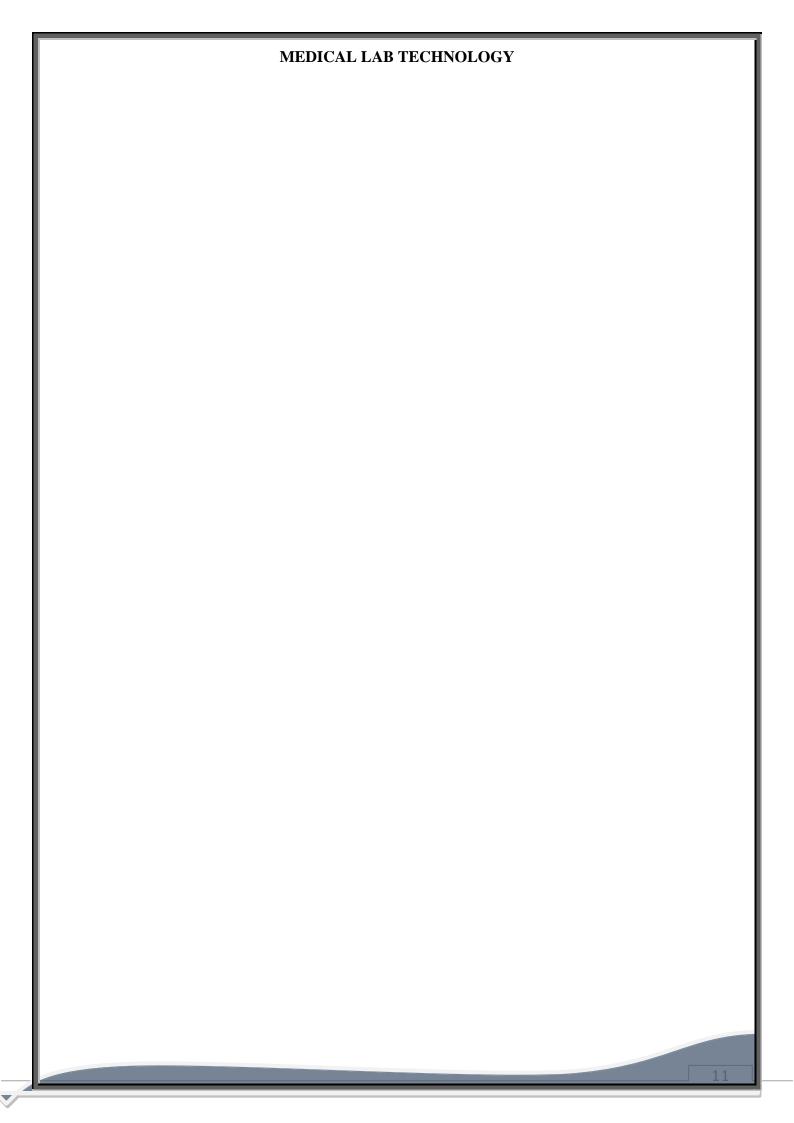
Marks allotted for evaluation: 100

S.No	Name of the activity	Max. Marks allotted for
		each activity
1	Biometric Attendance and punctuality	30
2	Familiarity with technical terms	05
3	Familiarity with tools and material	05
4	Manual skills	05
5	Application of knowledge	10
6	Problem solving skills	10
7	Comprehension and observation	10
8	Human relations	05
9	Ability to communicate	10
10	Maintenance of logbook	10
	Total	100

NOTE: The On the Job Training programme mentioned is tentative. The spirit of On the Job training is to be maintained. The colleges are at liberty to conduct on the job training according to their local feasibility of institutions & industries. They may conduct the entire on the job training periods/hrs of 365 for 1<sup>st</sup> year and 450 hrs for 2<sup>nd</sup> year either by conducting classes in morning session and send the students for OJT in afternoon session or two days in week or weekly or monthly or by any mode which is feasible for both the college and the institution. However, the total assigned periods/hrs for on the job training should be completed before 31<sup>st</sup> December

#### SCHEME OF INSTRUCTION PER WEEK

	Part-A	Theory	Practicals	Total
1.	English	4	-	4
2.	General Foundation Course	4	-	4
	Part-B			
3.	Paper –I	4	4	8
4.	Paper-II	4	4	8
5.	Paper-III	4	4	8
	Total	20	12	32



#### MEDICAL LAB TECHNOLOGY FIRST YEAR

#### PART B – VOCATIONAL SUBJECTS PAPER – I: BIO - CHEMISTRY (THEORY)

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
1.	Iı	ntroduction and Scope of Biochemistry	10	2	1	0
_,		thics and Disciplines		_	_	
		ab Ethics, Lab Discipline				
		mportance of Patient and Patients				
		•				
		Inagement for Medical Lab Technicians				
	aı	nd Medical Lab Organizations				
2.		eption, Identification, Registration of Chemical parameters for investigation	10	2	1	0
3.		sware and plastic ware used in bio-	08	6	0	1
		nical laboratory				
	I.	Glass ware:				
	a)	Types of glassware, their identification,				
	( <i>u</i> )	applications and uses.				
	b)	Cleaning, drying, maintenance and				
	U)					
	TT	storage of glassware.  Plastic ware: Brief outline.				
	II.	I .	00		0	1
4.		uments used in Bio-Chemistry Lab:	08	6	0	1
	I.	Colorimetry:				
		Visual and photoelectric methods,				
		Design (parts), principles and laws				
		involved construction, operation,				
		applications, care and maintenance.				
	II.	Spectrophotometry:				
		Principle, theory, types, construction,				
		operation, applications and Maintenance.				
5.	Basi	c Lab Operations	08	6	0	1
	I.	Separation of solids from liquids by				
	a)	Centrifugation: Principle, Different				
		types of centrifuges, construction of				
		electrical centrifuge, Procedure, care,				
		maintenance and applications.				
	b)	Filtration – using funnel.				
	II.	Weighing: Different types of balances –				
		simple balance, electronic balance –				
		operation, uses, care and maintenance.				
	III.	Evaporation				
	IV.	Distillation				
	V.	Refluxing				
	V.					
	V 1.	Drying different salts.	00	2	1	0
6.		Water Different Types, Chemicals and	08	2	1	0
	т	related substances				
	I.	Purity of chemicals				
	II.	Corrosives.				
	III.	hygroscopic substances				
7.		Hazards and Safety	8	6	0	1

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
		Physical, Chemical and Biological Hazards Awareness responsibility and measures of safety Prevention, communication and				
		Control of Lab Hazards				
8.		Collection of specimens	08	8	1	1
	I.	Blood – Phlebotomy [ How to draw the blood and precautions], types of Specimens, processing and preservation.				
	II.	Urine: Types of Specimens, Collection				
9.		of 24 hours urine – Preservatives.  Urine biochemical parameters.	08	2	1	0
10.		Units of measurements.	08	2	1	0
11.		Solutions	08	8	1	1
		Types based on solute and solvent, Types based on method of expressing concentration, Percentage calculations.			_	_
12.		Carbohydrates	04	6	0	1
		Carbohydrates- Definition, Biological importance, classification, qualitative tests.				
		Lipids: Definition, Biological importance, Classification and clinical importance.	4			
13.		Proteins and Amino Acids	08	2	1	0
		Definition, Biological importance of Proteins and Amino Acids Classification, Qualitative tests.				
14.		Diagnostic Tests: Blood Sugar- [GOD – POD Method ] Blood urea [ DAM-TSC Method, Enzymatic Method] Glucose tolerance Test [ GTT] Serum Uric Acid Serum Creatinine [ Jaffe's method /Alkaline picrate method]	19	8	1	1
15.		Vitamins:	04	2	1	0
		Vitamins classification Fat soluble vitamins, Water Soluble Vitamins, Sources, Daily requirements, Deficiency diseases.				
		Minerals: Sources, Daily requirements, Deficiency diseases.	04			
		Total	135	68	10	8

# MEDICAL LAB TECHNOLOGY FIRST YEAR PART B – VOCATIONAL SUBJECTS

PAPER – I: BIO - CHEMISTRY [PRACTICALS]

S.No.	Name of the Unit.	No. Of	Weightage
		Periods	in marks
1.	Reception and registration	10	3
2.	Collection of capillary blood	10	3
3.	Collection of venous blood.	10	4
4.	Collection of arterial blood.	10	4
5.	Separation of Serum and plasma from blood.	10	4
6.	Preparation of protein free blood filtrate.	10	4
7.	Lab glass ware	10	4
	a) Identification		
	b) Handling		
	c) Care and Maintenance		
	d) Uses.		
8.	Lab instruments	10	4
	a) centrifuges		
	b) Balances		
	c) Photo Electric Colorimeter		
	d) Spectrophotometer		
9.	Preparation of	10	4
	a) Percentage solutions		
	b) Normal Solutions		
	c) Molar Solutions.		
10.	Qualitative identification tests for sugars	10	4
11.	Qualitative identification tests for proteins.	10	4
12.	Quantitative determination of Blood Sugar	10	3
13.	Semi Quantitative determination of Urine Sugar.	15	5
	Total	135	50

#### MEDICAL LAB TECHNOLOGY FIRST YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II: MICROBIOLOGY & PATHOLOGY [THEORY]

S.No		PAPER – II: MICROBIOLOGY  NAME OF THE UNIT	No. Of	Weightage	Short	Essay/
			Periods	in marks	answer	Problem
					questions	questions
1.		oduction and Scope of Biochemistry	10	2	1	0
		es and Disciplines				
		Ethics, Lab Discipline				
	Impo	ortance of Patient and Patients				
	Man	agement for Medical Lab Technicians and				
	Med	ical Lab Organizations				
2.		eption, Identification, Registration of	10	2	1	0
		Chemical parameters for investigation				
3.		sware and plastic ware used in bio-	08	6	0	1
		nical laboratory				
	I.	Glass ware:				
	a)	Types of glassware, their identification,				
	1.	applications and uses.				
	b)	Cleaning, drying, maintenance and				
	**	storage of glassware.				
	II.	Plastic ware: Brief outline.	00			
4.		uments used in Bio-Chemistry Lab:	08	6	0	1
	I.	Colorimetry:				
		Visual and photoelectric methods,				
		Design (parts), principles and laws				
		involved construction, operation,				
	TT	applications, care and maintenance.				
	II.	Spectrophotometry:				
		Principle, theory, types, construction,				
	D 2	operation, applications and Maintenance.	00	(	0	1
5.		c Lab Operations	08	6	0	1
	I.	Separation of solids from liquids by				
	a)	Centrifugation: Principle, Different				
		types of centrifuges, construction of electrical centrifuge, Procedure, care,				
		maintenance and applications.				
	b)	Filtration – using funnel.				
	II.	Weighing: Different types of balances –				
	11.	simple balance, electronic balance –				
		operation, uses, care and maintenance.				
	III.	Evaporation				
	IV.	Distillation				
	V.	Refluxing				
	VI.	Drying different salts.				
6.	7 1.	Water Different Types, Chemicals and	08	2	1	0
0.		related substances			1	U
	I.	Purity of chemicals				
	II.	Corrosives.				
	III.	hygroscopic substances				
		75				
		77 7 7 7 7 7			0	4
7.		Hazards and Safety     Physical, Chemical and Biological	8	6	0	1

S.No	NAME OF THE UNIT		No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
		Hazards			•	•
		Awareness responsibility and				
		measures of safety				
		Prevention, communication and				
		Control of Lab Hazards				
		Control of Zuc Hubarus				
8.		Collection of specimens	08	8	1	1
	I.	Blood – Phlebotomy [ How to draw the				
		blood and precautions], types of				
		Specimens, processing and preservation.				
	II.	Urine: Types of Specimens, Collection				
		of 24 hours urine – Preservatives.				
9.		Urine biochemical parameters.	08	2	1	0
10.		Units of measurements.	08	2	1	0
11.		Solutions	08	8	1	1
		Types based on solute and solvent, Types				
		based on method of expressing				
		concentration, Percentage calculations.				
12.		Carbohydrates	04	6	0	1
		Carbohydrates-				
		Definition, Biological importance,				
		classification, qualitative tests.				
		Lipids:	4			
		Definition, Biological importance,				
10		Classification and clinical importance.	00			
13.		Proteins and Amino Acids	08	2	1	0
		Definition, Biological importance of				
		Proteins and Amino Acids Classification,				
1.1		Qualitative tests.	10	o	1	1
14.		Diagnostic Tests:	19	8	1	1
		Blood Sugar- [GOD – POD Method] Blood urea [ DAM-TSC Method,				
		Enzymatic Method]				
		Glucose tolerance Test [ GTT]				
		Serum Uric Acid				
		Serum Creatinine [ Jaffe's method				
		/Alkaline picrate method]				
15.		Vitamins:	04	2	1	0
		Vitamins classification		_	1	, , ,
		Fat soluble vitamins, Water Soluble				
		Vitamins, Sources, Daily requirements,				
		Deficiency diseases.				
		Minerals:	04			
		Sources, Daily requirements, Deficiency				
		diseases.				
		Total	135	68	10	8

#### MEDICAL LAB TECHNOLOGY FIRST YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II : MICROBIOLOGY & PATHOLOGY [PRACTICAL]

S.No	Name of the Unit	No. of Periods	Weightage in marks
	MICROBIOLOGY		
I	Lab instructions for personal safety precautions Good Laboratory Practices.	10	3
II	Receiving, Rejection criteria of a specimen, Registration	10	3
III	Cleaning of glassware (Old and New), and maintenance of lab equipment	5	2
IV.	Maintenance of Lab Equipment		
	Centrifuge, Incubator, Bio Safety Cabinet, Autoclave, Hot air Oven, Water bath		
V.	Handling and care of Microscope.	10	3
VI.	Operation of Autoclave, incubator, water bath, Hot air oven, Deep-freezer, centrifuge etc.,	10	3
VII.	Preparation of various media and biochemical sugars.	10	3
VIII.	Preparation of stains and smears.	10	4
IX	Quality control IQC and EQAS	10	4
X	Culture Techniques and inoculation methods		
XI	Bio Medical Waste Management		
	Syllabus for OJT same as above.		
I	PATHOLOGY		
II	Collection of blood	10	5
	- Receipt of requisition forms		
	- Receipt of samples		
	- Labeling		
III	Universal precautions	5	2
IV	Preparation of anticoagulants	08	3
V.	RBC, WBC & Platelet count.	08	3
VI.	ESR stands & ESR estimation.	08	3
VII	PCV – estimation.	5	2
VIII	Hb estimation by different methods.	08	3
	Urine- physical examination, Chemical examination & Microscopic	08	4
	examination.		
	- collection		
	- Preservatives		
	- Strip method.	46-	
	Total	135	50

#### MEDICAL LAB TECHNOLOGY I YEAR

# PART B – VOCATIONAL SUBJECTS

#### PAPER – III: ANATOMY & PHYSIOLOGY [THEORY]

S.No.	NAME OF THE			_	-
5.No.	NAME OF THE	No. Of	Weightage	Short	Essay/
	UNIT	Periods	in marks	answer	Problem
				questions	questions
I & II	Basics in Human				
	Anatomy &				
	Physiology				
1 & 13.	Introduction to Human	6+6	2	1	0
	Anatomy.				
	Introduction to Human				
	Physiology				
2 & 14.	Cell – Definition,	6+6+6	8	1	1
	Structure and				
	properties.				
	Tissue – Classification				
	in brief [ epithelial,				
	connective, muscular,				
	nervous]				
	Muscle physiology &				
	Nerve physiology				
3 & 16.	Respiratory system	6+5	8	1	1
4 & 17.	Digestive system &	6+6	8	1	1
	Hepato Biliary system				
5 & 18.	Cardio-vascular system.	5+5	8	1	1
6 & 15.	Lymphatic system.	5+5	2	1	0
	Blood & Lymph				
7.	Bones & Joints.	5	6	0	1
8, 12 & 22.	Nervous system – CNS	5+5+10	8	1	1
	Sense organs- Eye, ear,				
	Skin, nose, tongue				
	Nervous system [ CNS]				
	& Sense organs [ Eye,				
	ear, skin, nose, tongue]				
9 & 19.	Excretory system –	5+5	8	1	1
	Urinary system				
10 &20.	Endocrine system	5+9	8	1	1
11 & 21.	Reproductive system-	5+10	2	1	0
	Male & Female				
	Total	135	68	10	8

#### MEDICAL LAB TECHNOLOGY

#### I YEAR

#### PART B – VOCATIONAL SUBJECTS

### PAPER – III: ANATOMY & PHYSIOLOGY [PRACTICAL]

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks
I	Human Skeleton	TCHOOS	minarks
1.	Name of the Bones- Identification points, Surfaces of [Skull, Scapula, clavicle, humerus, radius, ulna, carpal bones, meta carpal bones, phalanges – Innominate bone, Femur, patella, tibia, fibula, tarsal bones, meta tarsal bones, Phalanges, Ribs-classification, vertebrae, sternum	30	10
2.	Human Organs[POP Models ]	20	09
	Brain, Stomach, Lungs, Intestines, Heart, Kidney, Liver, Uterus, Spleen, Fallopian tubes.		
3.	Human Slides [ Permanent Slides]	30	10
	Epithelial Tissue. Connective Tissue. Muscular Tissue. Nervous Tissue. Liver Kidney Spleen Pancreas Lymph nodes Skin Testes Ovary Uterus Tonsil Stomach layers Small intestine Large intestine.		
4.	Blood pressure	15	5
	Estimation of Blood pressure		
5.	T.P.R. [ Temperature, pulse, respiration ] chart	10	05
6.	TC,DLC [ RBC Total Count, WBC Total Count, differential Leucocytes count ]	30	11
	Total	135	50

#### MEDICAL LAB TECHNOLOGY SECOND YEAR

#### PART B – VOCATIONAL SUBJECTS PAPER – I : BIO- CHEMISTRY [THEORY]

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
I		strumental methods of Bio-Chemical alysis	10	6	0	1
	1.	Flame photometry: Principle, Theory, Construction of Flame Photometer, General and Clinical applications, study of electrolytes using flame photometer, clinical importance of determination of electrolytes.  New Instrument Automated Electrolyte				
	2.	Analyzer  Fluorometry: Definition Principle, Theory and Construction of Fluorometer and its clinical Applications, general and Clinical applications.				
II		Separation Techniques.	10	8	1	1
	2	Electrophoresis: Definition and basic principle, Procedure, Different types, and Clinical Applications, Example:-Electrophoretic fractionation of serum protein and Serum lipo proteins.  Chromatography: Definition, Basic				
		Principles, Different types, Procedure and Clinical applications.				
III.		Immuno Assays – Definition, Basic Principles of Immuno chemical reactions and immune assays.	10	2	1	0
	1.	Radio immune assays.  Introduction to radioactivity, Radioactive substances and its applications Ex:- Thyroid Hormone Assays  [T3,T4,TSH]				
	2.	Enzyme linked immune sorbent assays (ELISA). Description, Instruments used in these assays, Applications				
IV.		Metabolism:	10	6	0	1
	1.	Carbohydrate metabolism – Glycolysis,				
	2.	TCA Cycle and Gluconeogenesis.  Lipid metabolism- β-Oxidation of Fatty Acids.				
	3.	Protein metabolism – Urea cycle.				
V.	· ·	Titrimetric methods of quantitative determination, preparation of various solutions used in Titrimetric analysis.	5	2	1	0
		Organ Function Tests				
VI		Liver Function Tests[ LFT]: Functions of Liver	10	8	1	1

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
		Metabolism of Hb. Bilirubin Explanation Jaundice and its types Determination of Serum Bilirubin(Vandenberg's Tests)			questions	questions
	4.	Determination of total Serum proteins (Albumin and Globulin) & A/G Ratio.				
	5.	Estimation of Enzymes – SGOT, SGPT, Alkaline Phosphatase (ALP) & Acid phosphatase				
VII.		Kidney function tests [KFT/RFT]: Functions of Kidney Synthesis of Urea Basic Concepts Estimation of Blood Urea DAM-TSC Method Estimation of Serum Creatinine [Jaffe's Method / Alkaline Picrate] Clearance Test - Definition Creatinine Clearance, Urea Clearance Urine Examination in accessing KFT Abnormal Constituents of Urine (Sugar Ketone bodies, Protein Blood bile salts and bile pigments)	10	8	1	1
VIII.		Gastric Function Tests [ GFT]	5	2	1	0
	1.	Introduction & Basic concepts.				
	2.	Estimation of Free HCL combined Acids – Clinical significance.				
IX.		Thyroid Function Tests [ TFT]	5	2	1	0
	1.	Introduction - Thyroid Gland and its Functions			_	
	2.	Estimation of Thyroid Hormones [ T3,T4,TSH] - Methods – RIA &Chemiluminiscence and its Clinical significance.				
X.		Pancreatic Function Test [ PFT]	5	2	1	0
	1.	Introduction of Basic concepts.				
XI.	2.	Determination of Serum amylase.  Clinical Enzymology:	5	6	0	1
AI.	1.	Introduction & Basic concepts of Enzymes, Co-Enzymes, Iso Enzymes.	3	U	U	1
	2.	Importance of Enzymes.				
	4	Liver Enzymes - SGPT, ALP and GGT Cardiac Enzymes - CPK, LDH, SGOT.				
	5.	Acid Phosphatase.				
XII.		<b>Body Fluids:</b>	5	2	1	0
	1. 2.	Outlines of formations of different body fluids  Composition & Analysis of CSF				
		including.				

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	<ul><li>a) CSF Sugar estimation</li><li>b) CSF Proteins estimation</li><li>c) CSF Chlorides estimation.</li><li>Including interpretation of results.</li></ul>				
XIII.	Automation of Biochemistry Labs and usage of Computers.	5	6	0	1
XIV	Quality Control	5	2	1	0
	<ul><li>a) Introduction and importance of quality assurance, General principle.</li><li>b) Internal and external quality control.</li></ul>				
XV	Diagnostic Tests :	10	6	0	1
	<ul> <li>Blood Glucose</li> <li>Blood Urea</li> <li>Serum Creatinine</li> <li>Plasma Proteins</li> <li>Lipid Profile - Serum Total cholesterol, Tri Glycerides HDL Cholesterol, LDL cholesterol and VLDL Cholestrol.</li> <li>GlycoselatedHaemoglobin</li> <li>Serum Calcium and Inorgonic Phosphate</li> <li>Arterial Blood Gas Analysis [PH, PO2 AND PCO2]</li> </ul>				
	[111,102711011002]	110	68	10	8

# MEDICAL LAB TECHNOLOGY SECOND YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – I : BIO- CHEMISTRY [PRACTICALS]

S.No.	Name of the Unit	No. Of Periods	Weightage in marks
I.	BLOOD TESTS		
	Estimation of Blood Glucose	10	10
	Estimation of Blood Urea		
	Estimation of Serum Creatinine		
	Plasma Proteins		
	Estimation of Serum Cholestrol		
	Lipid Profile		
	Enzymes		
	a) SGOT		
	b) SGPT		
	c) ALP		
	Vandenberg's Test (Estimation of Bilirubin)		
	Estimation of Calcium and Inorganic Phosphates		
	Estimation of Serum Electrolytes		
	URINE TESTS	5	6
	Identification of Ab normak constituents of Urine		

	( Sugar, Ketone Bodies, Protein, Blood, Bilesalts and		
	Bilepigments)		
	DEMONSTRATION	5	3
	Oral Glucose Tolerance Test(OGTT)		
	Electrophorosis		
	Chromatography		
	CSF ANALYSIS	3	2
	Bio Chemical Analysis of Sugar and Proteins		
1.	Electrophoretic fractionation of serum proteins and lipo proteins –	3	2
	Demo.		
2.	Separation of amino acids and carbohydrates by paper	3	2
	chromatography – Demo.		
3.	Determination of plasma prothrombin time	2	2
4.	Oral glucose tolerance test [GTT]- Demo	2	2
5.	Estimation of serum calcium and inorganic phosphate.	2	2
6.	Practice and use of automated pipettes.	5	2
7.	Estimation of HDL cholesterol.	5	2
8.	Determination of urine proteins by turbidmetric method – [Sulfosalysilic Acid ]	5	2
9.	CSF analysis – Pandy's test Nonne-Apelt – Sugars, Proteins	5	2
10.	Demonstration of working of Auto analysers.	10	5
11.	Training of Computer basics.	15	7
12.	Estimation of serum sodium and potassium by Flame photometry.	5	3
13.	Qualitative identification of urine sugars.	5	2
14.	Qualitative identification of urine proteins [ Heat Coagulations}	5	2
15.	Qualitative identification of urine Bile Salts & Bile Pigments.	5	2
16.	Determination of Serum Bilirubin, SGPT & Alkaline Phosphatase [LFT]	10	5
17.	Determination of Blood Urea and serum creatinine [KFT]	10	4
18.	Practice and interpretation of lipid profile.	5	2
	Total	115	50

# MEDICAL LAB TECHNOLOGY SECONDYEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II: MICROBIOLOGY [THEORY]

S.N o	NAME OF THE UNIT	No. Of Periods	Weight age in marks	Short answer questio ns	Essay/ Proble m question s
I	Normal Flora of Micro-organisms in the Human Body	5	2	1	0
II	Introduction to Immunology:	10	8	1	1
a)	Brief outline of Immunity				
b)	What are antigens?				
c)	What are antibodies?				
d)	Different types of antigen and antibody reactions, their applications in the diagnostics - agglutination, precipitation, complement fixation, Neutralisation, RIA.				
e)	Principle and method of ELISA Test.	4.0			
III	Collection and processing of faecal samples, concentration techniques of stool for Microscopic Examination	10	8	1	1
	Parasitology:				
IV.	Antibiotic sensitivity Test – preparation of Antibiotic discs.	5	6	0	1
V.	Preservation methods of stock cultures and their importance and principle - procedure	5	2	1	0
VI.	Brief outline of Morphology cultural characteristics and Lab diagnosis of imp. Pathogens.	25	14	1	2
a)	Gram Positive – Staphylococcus, Streptococcus, Pneumococcus				
b)	Gram Negative cocci– Gonococci, Meningococci.				
c)	Gram Positive Bacilli- Corynebacterium  – diphtheriae, Mycobacterium tuberculosis, Mycobacterium leprae				

S.N o	NAME OF THE UNIT	No. Of Periods	Weight age in marks	Short answer questio ns	Essay/ Proble m question s
d)	Gram Negative Bacilli –				
	Enterobacteriaceae – E.coli, Klebsiella,				
	Salmonella, Shigella.				
e)	Anaerobic Bacteria- Bacteriodes,				
	Clostridium spp.				
f)	Vibriocholera, Pseudomonas.				
g)	H.influenza, B.pertusis.				
<u>h)</u>	Spirochetes- Treponema, Leptospira.				
i)	Actinomyces&Nocardia.	10		-	
VII.	Bacteriological Examination of Water, Milk & Food.	10	8	1	1
VIII	Mycology	15	8	1	1
•	Morphology, cultural characteristics and lab diagnosis of :				
IX	Virology	10	8	1	1
	Classification, General properties and cultivation of imp. pathogenic viruses such as Polio, Hepatitis, Rabies, HIV and Dengue.				
X.	Quality Control in Laboratory.	10	2	1	0
XI	Automation in Clinical Laboratories - in brief.	5	2	1	0
	Total	110	68	10	8

# MEDICAL LAB TECHNOLOGY SECOND YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II: MICROBIOLOGY [PRACTICAL]

S.No	NAME OF THE UNIT	No. Of	Weightage
		Periods	in marks
1.	Parasitology	10	5
	a) Collection, preservation and transportation of faecal		
	material for examination of parasites.		
	b) Concentration techniques of stool for ova and cysts.		
	c) Wet preparation of faecal sample for ova and cysts.		
2.	Procedure, processing of sputum for AFB	5	2
3.	Procedure of skin clipping of leprae bacilli.	5	2
4.	Inoculation techniques on media and putting up biochemical	10	4
	reactions for the isolation of common organisms like –		
	Staphylococcus, E.coli, Klebsiella, Shigella, Salmonella,		
	Proteus, Pseudomonas, Automated Identification Systems		
5.	Preparation of antibiotic discs and putting Sensitivity Tests,	10	6
	Automated Sensivity Testing Systems.		
6.	Preservation and Maintenance of Stock Cultures.	5	2
7.	Collection and processing of Clinical Samples for Culture.	15	5
	a) Blood		
	<ul> <li>Collection of Blood</li> </ul>		
	<ul> <li>Blood Culture Media</li> </ul>		
	<ul> <li>Incubation</li> </ul>		
	<ul> <li>Direct Staining</li> </ul>		
	<ul> <li>Subculture and Identification</li> </ul>		
	<ul> <li>Automated Bed Culture System</li> </ul>		
	b) Urine		
	<ul> <li>Sample Collection, Transport and Storage</li> </ul>		
	Media for inoculation		
	<ul> <li>Incubation</li> </ul>		
	Semi Quantitative Colony Count		
	c) Stool		
	Sample Collection, Transport and Storage		
	<ul> <li>Media – Transport, Enrichment, Selective</li> </ul>		
	Inoculation of Media		
	<ul><li>Identification of Pathogens</li><li>d) Sputum</li></ul>		
	•		
	• Sample Collection		
	Adequacy of Specimen     Inaculation of Madia		
	Inoculation of Media		
	Identification of Pathogens     Francisco Production Producti		
	e) Exudates – Pus / Throat Swab, Vaginal Swab etc.,		
	• Sample Collection		
	• Direct Smear		
	<ul> <li>Inoculation of Media</li> </ul>		
	<ul> <li>Identification of Pathogens</li> </ul>		

	f) Sterile Fluids		
	• CSF		
	<ul> <li>Centrifugation</li> </ul>		
	Direct Smear		
	<ul> <li>Inoculation of Media</li> </ul>		
	<ul> <li>Identification of Pathogens</li> </ul>		
	g) Other Fluids – Peritoneal, Pleural, Liver Aspirates		
	Sample Collection		
	Centrifugation		
	Direct Smears		
	<ul> <li>Inoculation of Media</li> </ul>		
	<ul> <li>Identification of Pathogens</li> </ul>		
8.	Collection of specimen for fungal examination like skin	5	2
	scrapings, CSF & Nail clippings.		
9.	Processing like germ tube tests ,culture of samples, LCB	5	2
	Mounts		
	Serology		
10.	CRP, ASO, RA, VDRL, Widal, Brucella, ELISA, Western	35	16
	blot tests.		
	Virology		
11.	<ul> <li>Incubation of fertile eggs and inoculation by various</li> </ul>	5	2
	routes		
	Collection of Swabs for Swine flu		
12.	Laboratory Information System	5	2
	Total	115	50

**Syllabus for OJT same as above.** 

#### MEDICAL LAB TECHNOLOGY SECOND YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – III: PATHOLOGY [THEORY]

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
I	Preparation of blood smears and examination:  - Thin & thick blood films.  - Making an ideal blood film.  - Methods – slide method, Cover slip method  - Staining – composition, preparation & procedure of leishman stain.  - Knowledge about Romanowsky stains.  - Examination – Morphology & Identification of RBC, WBC & Platelets  - Counting – DLC – Counting methods, Normal values, clinical significance and limitations.  - Oils used for immersion- types	08	8	1	1
II	Special stains for Bone marrow smears:  - Giemsa, Wrights, Myeloperoxidase stain, Periodic Acid Schiff [PAS]- Composition, Preparation, procedure and interpretation.	5	2	1	0
III	Bone Marrow Aspiration / trephine biopsy:  - Setting up of tray for bone marrow aspiration - Preparing smears – methods- Imprints, crush Staining, clinical significance.	5	2	1	0
IV	Identification of hemoparasites:  - Morphology of malaria parasite, microfilaria, leishmania, trypanosomiasis.  - Importance of sample collection time.  - Making thick and thin smears.  - Procedure of making & staining the smears.  - Identification of the parasite.	5	6	0	1
V	Absolute Eosinophil count:  - Materials required, diluting fluids, procedure, and identification and counting of cells.	5	2	1	0
VI	Sickle cell preparation: - Principle, procedure, methods, Materials required, clinical significance.	5	6	0	1
VII	Osmotic fragility test:  - Methods used, materials required, procedure, observation, reporting, Normal values, factors affecting and interpretation.	5	2	1	0
VIII	Coagulation Tests:  a) Bleeding time- methods- Dukes method, lvy's method – procedure, normal values and clinical significance.  b) Clotting time – methods- Lee & White, capillary tube method- procedure,	08	6	0	1

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	materials, normal values, factors affecting coagulation and clinical significance. c) Prothrombin time [PT] d) APTT e) Introduction to Coagulometers			questions	questions
IX	Buffy coat preparation : L.E. cells, microfilaria and abnormal cells.	5	2	1	0
X. *	L.E cell Test: - Principle, procedure, material required, reporting, clinical significance.	5	2	1	0
XI.	Basics of cell counter:  - Diluting fluids - Maintenance of counter Quality Control and is significance	5	2	1	0
XII.	Histopathology:  - Maintenance of Registers – receiving register, gross register, Slide register and report issue register.  - Biopsy & tissue specimens – Example: Thyroid, GIT, breast, soft tissue, bone etc.  - Fixatives - processing - Dehydration - clearing - Impregnation - paraffin embedding and block making - Trimming of blocks Tissue cutting [ Microtomes] - Staining of the sections Mounting & Mounting Media, cover slips, labeling Decalcification of bone & calcified tissue Routine Hematoxylin& Eosin staining Immunohistochemistry Special stains- PAS, Reticulin, Perls, Masson's Trichrome etc.	13	6	0	1
XIII.	<ul> <li>Filing of slides, storing of blocks etc.</li> <li>Microtomes &amp; Knives:         <ul> <li>Types of Microtomes – Maintenance.</li> <li>Sharpening of Knives – Honing &amp; Stropping</li> <li>Disposable Blades – Types and Care</li> <li>Advantages and dis-advantages of frozen section cutting.</li> </ul> </li> </ul>	08	2	1	0
XIV	Cytology: - FNAC - Guided aspiration - Pap smear - Cytospin – equipment – machine, Procedure, material, laying a tray for the procedure Making smears, staining & mounting cover slip, labeling Identification & Interpretation – basics.	08	6	0	1
XV	Museum techniques:	5	6	0	1

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	<ul> <li>Labeling &amp; storage of specimens</li> </ul>				
	<ul> <li>Methods of color maintenance.</li> </ul>				
	- Presentation of specimen.				
	<ul> <li>Mounting, Labeling and cataloging the</li> </ul>				
	specimen				
	<ul> <li>Maintenance and cleanliness of the</li> </ul>				
	Museum.				
	- Disposal of waste,				
	- Safety in the lab.				
XVI	Autopsy:	5	2	1	0
	-Aims & methods of performing Autopsy,				
	cleaning, suturing and restoring the body.				
	Cleaning the autopsy instruments, tables and				
	rooms, preservation of organs.				
XVII.	ImmunoHematology and Blood Banking:	10	6	0	1
	- Introduction				
	- Human blood group antigens, their				
	inheritance and antibodies.				
	- ABO Blood group systems.				
	- RH Blood group system.				
	<ul> <li>Techniques of grouping and cross</li> </ul>				
	matching.				
	- Blood collection, screening of donor,				
	preservation and maintenance of records.				
	- Coombs Test – a) Direct b) indirect.				
	Total	110	68	10	8

#### MEDICAL LAB TECHNOLOGY SECOND YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – I : BIO- CHEMISTRY [THEORY]

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
I		trumental methods of Bio- emical Analysis	10	8	1	1
	1.	Flame photometry: Principle, Theory, Construction of Flame Photometer, General and Clinical applications, study of electrolytes using flame photometer, clinical importance of determination of electrolytes. New Instrument Automated				
		Electrolyte Analyzer				
II		Separation Techniques.	10	8	1	1
	1.	Electrophoresis: Definition ,basic principle,and Clinical Applications, Example:- Electrophoretic fractionation of serum protein and Serum lipo proteins.				
	2	Chromatography: Definition, Basic Principles, Different types, Procedure and Clinical applications of Paper Chromatography				
III.		Immuno Assays – Definition and Basic Principle.	10	8	1	1
	1.	Radio immune assays. Introduction and Clinical Applications.				
	2.	Enzyme linked immune sorbent assays (ELISA).  Description, Instruments used in these assays, Applications				
IV.		Metabolism:	10	6		1
	1.	Carbohydrate metabolism – Glycolysis, TCA Cycle.				
	2.	Lipid metabolism Importance				
	3.	Protein metabolism – Importance				
V.		Titrimetric methods of quantitative determination. Concept of stock standard solutions, working standard solutions.	5			
		Organ Function Tests				
VI		Liver Function Tests[ LFT]: Determination of Serum Bilirubin(Vandenberg's Tests)	10	8	1	1

S.No		NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	4.	Determination of total Serum proteins (Albumin and			questions	questions
		Globulin) & A/G Ratio.				
	5.	Estimation of Enzymes –				
		SGOT, SGPT, Alkaline				
		Phosphatase (ALP) & Acid				
VII.		phosphatase  Kidney function tests [	10	8	1	1
V 11.		Kruney function tests [ KFT/RFT]:	10	ð	1	1
		Introduction and classification				
		Estimation of Blood Urea				
		DAM-TSC Method				
		Estimation of Serum				
		Creatinine				
		[Jaffe's Method / Alkaline				
		Picrate] Clearance Test - Definition				
		Creatinine Clearance, Urea				
		Clearance				
VIII.		Gastric Function Tests [ GFT]	5	2	1	
	1.	Introduction .				
	2.	Estimation of Free HCL				
		combined Acids – Clinical				
137		significance.	-	2	1	
IX.		Thyroid Function Tests [ TFT]	5	2	1	
	1.	Introduction - Thyroid Gland				
		and its Functions				
	2.	Estimation of Thyroid				
		Hormones [ T3,T4,TSH] -				
		Methods – RIA & Chemiluminiscence and its				
		Clinical significance.				
X.		Pancreatic Function Test [	5	2	1	
		PFT]		_	_	
	1.	Introduction of Basic concepts.				
	2.	Determination of Serum				
***		amylase.	_		4	
XI.	1	Clinical Enzymology:	5	6	1	
	1.	Introduction & Basic concepts of Enzymes, Co-Enzymes, Iso				
		Enzymes.				
	2.	Importance of Enzymes. In				
		Diagnosis of Liver and Cardiac				
		condition				
	3	Liver Enzymes - SGPT, ALP and GGT				
	4	Acid Phosphatase.				
		•				
XII.		Body Fluids:	5	2	1	
	1.	Outlines of different body fluids				
	2.	Analysis of CSF including.				

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	<ul> <li>d) CSF Sugar estimation</li> <li>e) CSF Proteins estimation</li> <li>f) CSF Chlorides estimation.</li> <li>Including interpretation of results.</li> </ul>				
XIII.	Automation of Biochemistry Labs and usage of Computers in Medical field	5	6	1	
XIV	Quality Control	5			
	c) Introduction and importance of quality assurance,Internal and external quality control.				
XV	Diagnostic Tests :	10	8	1	
	<ul> <li>Blood Glucose</li> <li>Lipid Profile - Serum Total cholesterol, Tri Glycerides HDL Cholesterol, LDL cholesterol and VLDL Cholestrol.</li> <li>Glycoselated Haemoglobin</li> <li>Serum Calcium and Inorgonic Phosphate</li> </ul>				
		110			1

# MEDICAL LAB TECHNOLOGY SECONDYEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II: MICROBIOLOGY [THEORY]

S.N o	NAME OF THE UNIT	No. Of Periods	Weight age in marks	Short answer questio ns	Essay/ Proble m question
I	Normal Flora of Micro-organisms in the Human Body	5	2	1	
II	Introduction to Immunology:	10	8	1	1
a)	Brief outline of Immunity				
b)	What are antigens?				
c)	What are antibodies?				
d)	Different types of antigen and antibody reactions, their applications in the diagnostics - agglutination, precipitation, complement fixation, Neutralisation, RIA.				
e)	Principle and method of ELISA Test.		_	_	_
III	Collection and processing of faecal samples, concentration techniques of stool for Microscopic Examination	10	8	1	1
	Parasitology:				
IV.	Antibiotic sensitivity Test – preparation of Antibiotic discs.	5	6		1
V.	Preservation methods of stock cultures and their importance and principle procedure.	5	4	2	
VI.	Brief outline of Morphology cultural characteristics and Lab diagnosis of imp. Pathogens.	25	16	2	2
a)	Gram Positive – Staphylococcus, Streptococcus, Pneumococcus				
b)	Gram Negative cocci– Gonococci, Meningococci.				
c)	Gram Positive Bacilli- Corynebacterium – diphtheriae, Mycobacterium tuberculosis, Mycobacterium leprae				
d)	Gram Negative Bacilli – Enterobacteriaceae – E.coli, Klebsiella, Salmonella, Shigella.				
e)	Anaerobic Bacteria- Bacteriodes,				

S.N o	NAME OF THE UNIT	No. Of Periods	Weight age in marks	Short answer questio ns	Essay/ Proble m question s
	Clostridium spp.				
f)	Vibriocholera,.				
g)	H.influenza, B.pertusis.				
h)	Spirochetes- Treponema.				
i)	Actinomyces & Nocardia.				
VII.	Bacteriological Examination of Water,	10	10	2	1
	Milk & Food.				
VIII	Mycology.	15	12	3	1
•					
	Morphology, cultural characteristics and				
	lab diagnosis of :				
	Candida, Cryptococcus,				
	Dermatophytes, Aspergillus,				
	Penicillum.				
IX	Virology	10	8	1	1
	Classification, General properties and				
	cultivation of imp.pathogenic viruses				
	such as Polio, Hepatitis, Rabies, HIV				
	and Dengue.				
X.	Quality Control in Laboratory.	10	8	1	1
XI	Automation in Clinical Laboratories -	5	6		1
	in brief.				
	Total	110			

#### MEDICAL LAB TECHNOLOGY SECOND YEAR

# PART B – VOCATIONAL SUBJECTS PAPER – II : MICROBIOLOGY [PRACTICALS]

S.No	NAME OF THE UNIT	No. Of	Weightage
		Periods	in marks
1.	Parasitology	10	5
	d) Collection, preservation and transportation of faecal		
	material for examination of parasites.		
	e) Concentration techniques of stool for ova and cysts.		
	f) Wet preparation of faecal sample for ova and cysts.	_	
2.	Procedure, processing of sputum for AFB	5	2
3.	Procedure of skin clipping of leprae bacilli.	5	2
4.	Inoculation techniques on media and putting up biochemical	10	4
	reactions for the isolation of common organisms like –		
	Staphylococcus, E.coli, Klebsiella, Shigella, Salmonella,		
	Proteus, Pseudomonas, Automated Identification Systems	1.0	
5.	Preparation of antibiotic discs and putting Sensitivity Tests,	10	6
	Automated Sensivity Testing Systems	_	
6.	Preservation and Maintenance of Stock Cultures.	5	2
7.	Collection and processing of Clinical Samples for Culture.	15	5
	h) Blood		
	<ul> <li>Collection of Blood</li> </ul>		
	<ul> <li>Blood Culture Media</li> </ul>		
	<ul><li>Incubation</li></ul>		
	<ul> <li>Direct Staining</li> </ul>		
	<ul> <li>Subculture and Identification</li> </ul>		
	<ul> <li>Automated Bed Culture System</li> </ul>		
	i) Urine		
	<ul> <li>Sample Collection, Transport and Storage</li> </ul>		
	<ul> <li>Media for inoculation</li> </ul>		
	<ul><li>Incubation</li></ul>		
	<ul> <li>Semi Quantitative Colony Count</li> </ul>		
	j) Stool		
	<ul> <li>Sample Collection, Transport and Storage</li> </ul>		
	Media – Transport, Enrichment, Selective		
	Inoculation of Media		
	Identification of Pathogens		
	k) Sputum		
	Sample Collection		
	Adequacy of Specimen		
	Inoculation of Media		
	<ul> <li>Identification of Pathogens</li> </ul>		
	l) Exudates – Pus / Throat Swab, Vaginal Swab etc.,		
	_		
	• Sample Collection • Direct Smoor		
	Direct Smear     Inaculation of Modic		
	Inoculation of Media  In a Section 1.1.  In a		
	<ul> <li>Identification of Pathogens</li> </ul>		

8.	Collection of specimen for fungal examination like skin	5	2
	scrapings, CSF & Nail clippings.		
	Serology		
9.	CRP, ASO, RA, VDRL, Widal, ELISA, Western blot tests.	35	16
	Total	115	50

**Syllabus for OJT same as above.** 

## MEDICAL LAB TECHNOLOGY SECOND YEAR

## PART B – VOCATIONAL SUBJECTS PAPER – III: PATHOLOGY [THEORY]

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
I	Preparation of blood smears and examination:  - Thin & thick blood films.  - Making an ideal blood film.  - Methods – slide method, Cover slip method  - Staining – composition, preparation & procedure of leishman stain.  - Knowledge about Romanowsky stains.  - Examination – Morphology & Identification of RBC, WBC & Platelets  - Counting – DLC – Counting methods, Normal values, clinical significance and limitations.  - Oils used for immersion- types	08	8	1	1
II	Special stains for Bone marrow smears:  - Giemsa, Wrights, Myeloperoxidase stain, Periodic Acid Schiff [PAS]- Composition, Preparation, procedure and interpretation.	5	2	1	
III	Bone Marrow Aspiration / trephine biopsy:  - Setting up of tray for bone marrow aspiration - Preparing smears – methods- Imprints, crush Staining, clinical significance.	5	2	1	
IV	Identification of hemoparasites:  - Morphology of malaria parasite, microfilaria, leishmania, trypanosomiasis.  - Importance of sample collection time.  - Making thick and thin smears.  - Procedure of making & staining the smears.  - Identification of the parasite.	5	6		1
V	Absolute Eosinophil count:  - Materials required, diluting fluids, procedure, and identification and counting of cells.	5	2	1	
VI	Sickle cell preparation: - Principle, procedure, methods, Materials required, clinical significance.	5	6		1
VII	Osmotic fragility test:  - Methods used, materials required, procedure, observation, reporting, Normal values, factors affecting and interpretation.	5	2	1	
VIII	Coagulation Tests:  f) Bleeding time- methods- Dukes method, lvy's method – procedure, normal values and clinical significance.  g) Clotting time – methods- Lee & White, capillary tube method- procedure,	08	6		1

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	materials, normal values, factors affecting coagulation and clinical significance.  h) Prothrombin time [PT]  i) APTT			questions	questions
	j) Introduction to Coagulometers				
IX	Buffy coat preparation: L.E. cells, microfilaria and abnormal cells.	5	2	1	
X. *	L.E cell Test: - Principle, procedure, material required, reporting, clinical significance.	5	2	1	
XI.	Basics of cell counter:  - Diluting fluids - Maintenance of counter Quality Control and is significance	5	2	1	
XII.	Histopathology:  - Maintenance of Registers – receiving register, gross register, Slide register and report issue register.  - Biopsy & tissue specimens – Example: Thyroid, GIT, breast, soft tissue, bone etc.  - Fixatives - processing - Dehydration - clearing - Impregnation - paraffin embedding and block making - Trimming of blocks Tissue cutting [ Microtomes] - Staining of the sections Mounting & Mounting Media, cover slips, labeling Decalcification of bone & calcified tissue Routine Hematoxylin & Eosin staining Immuno histochemistry Special stains- PAS, Reticulin, Perls, Masson's Trichrome etc Filing of slides, storing of blocks etc.	13	16	2	2
XIII.	- Filing of slides, storing of blocks etc.  Microtomes & Knives:  - Types of Microtomes – Maintenance.  - Sharpening of Knives – Honing & Stropping  - Advantages and dis-advantages of frozen section cutting.	08	2	1	
XIV	Cytology: - FNAC - Pap smear - Cytospin – equipment – machine, - Procedure, material, laying a tray for the - procedure Making smears, staining & mounting - cover slip, labeling Identification & Interpretation – basics.	08	6		1
XV	Museum techniques: - Labeling & storage of specimens -	5	6		1

S.No	NAME OF THE UNIT	No. Of Periods	Weightage in marks	Short answer questions	Essay/ Problem questions
	- Mounting, Labeling and cataloging the				
	specimen				
	- Maintenance and cleanliness of the				
	Museum.				
	- Safety in the lab.	_			
XVI	Autopsy:	5	2	1	
	-Aims & methods of performing Autopsy,				
	cleaning, suturing and restoring the body.				
	Cleaning the autopsy instruments, tables and				
	rooms, preservation of organs.				
XVII.	Immuno Hematology and Blood Banking:	10	6		1
	- Introduction				
	<ul> <li>Human blood group antigens, their</li> </ul>				
	inheritance and antibodies.				
	<ul> <li>ABO Blood group systems.</li> </ul>				
	<ul> <li>RH Blood group system.</li> </ul>				
	<ul> <li>Techniques of grouping and cross</li> </ul>				
	matching.				
	- Blood collection, screening of donor,				
	preservation and maintenance of records.				
	<ul> <li>Coombs Test – a) Direct b) indirect.</li> </ul>				
	Total	110			

## MEDICAL LAB TECHNOLOGY II YEAR

## PART B – VOCATIONAL SUBJECTS PAPER – III:PATHOLOGY [PRACTICALS]

S.No	NAME OF THE UNIT	No. Of	Weightage
		Periods	in marks
I	1.Maintenance, Cleaning and care about Automatic tissue processor	15	7
	2. Microtomes & knives – sharpening		
II.	Maintenance & Preservation of cytology slides, histopathology	10	5
	blocks & slides – Histopathology specimens & Processing.		
	- Preparation of formalin for fixation.		
III.	Glass Ware	5	2
	- Slides, Cover slips, sample collection jars, test tubes and		
	watch glasses.		
IV.	Immuno Hematology & Blood Banking	20	8
	<ul> <li>ABO Blood grouping techniques</li> </ul>		
	- RH Factor		
	- Cross matching.		
	- Coombs test- Direct & indirect methods.		
V.	Histopathology:	20	8
	1. Fixation of Tissue		
	2. Processing		
	3. Dehydration		
	4. Clearing		
	5. Impregnation		
	6. Paraffin embedding and block making.		
	7. Trimming of block		
	8. Staining of the sections		
	9. Special stains – PAS, Perls, Reticulin and Masson's		
	Trichrome.		
	10. Mounting & labeling		
* * * *	11. Decalcification – to be told separately.	1.5	
VI.	Cytology:	15	7
	Fixatives.		
	Cytological exam of all fluids		
	Slide preparation, staining		
<b>T</b> / <b>T T</b>	Pap smears – staining, labelling, mounting & preservation	~	2
VII	Sickle Cell preparation	5	2
VIII	Buffy Coat preparation	5	2
IX	Bone marrow smears- preparation & staining.	5	2
X	Coagulation tests – B.T, C.T., P.T, A.P.T.T etc.,	15	7
	Total	115	50

**Example**: If the student strength is 40 they can attend by rotation as below. For first year

Outpatient Biochemistry Lab Reception, Sample collection Accepting and rejecting criteria for samples. Reception, Sample collection Accepting and rejecting criteria for samples. Reception, Sample collection Accepting and rejecting criteria for samples. Reception, Sample collection Accepting and rejecting criteria for samples. Reception, Sample collection and rejecting criteria for samples.  Main Lab Biochemistry Helping the senior technicians in cestimation of Various analytes 1. Preparing the anticoagulants 2. Preparing the Leishman's stain. 3. Preparing peripheral blood smear. 4. Staining of peripheral blood smear. 5. Preparing and staining thick smear 6. Estimation of haemoglobin by sahl's method and by photoelectric method. 7. Estimation of ESR 8. Estimation of PCV by Micro and Macro Methods. 9. Physical examination of Urine. 10. Chemical Examination of urine for sugars-manual method/strip method. 11. Chemical Examination of urine for Retone bodies-manual method/strip method. 12. Chemical Examination of urine for Protiens-manual method/strip method. 13. Chemical Examination of urine for Die pigments - manual method/strip method. 14. Chemical Examination of urine for bile pigments - manual method/strip method. 15. Chemical Examination of urine for bile pigments - manual method/strip method. 16. Preparing wet smear for urine microscopy. 17. Preparing processing the blood sample for reticulocyte count by wet and dry method. 18. Charging the improved Neubauer chamber	:If the student strength is 40 th	ey can attend by rotation as below.	
Accepting and rejecting criteria for samples.  Outpatient Pathology Lab Reception, Sample collection Accepting and rejecting criteria for samples.  Outpatient Microbiology Lab Reception, Sample collection amples.  Main Lab Biochemistry Reception, Sample collection criteria for samples.  Main Lab Biochemistry Reception, Sample collection are selected for samples.  Main Lab Pathology  I Preparing the senior technicians in estimation of Various analytes Preparing the Leishman's stain.  Preparing peripheral blood smear.  Seriam and staining thick smear.  Seriam and staining thick smear.  Ferparing and staining thick smear.  Physical examination of Lorine.  Chemical Examination of Urine.  Chemical Examination of urine for Sugars-manual method/strip method.  Chemical Examination of urine for Retone bodies-manual method/strip method.  Chemical Examination of urine for Protiens-manual method/strip method.  Chemical Examination of urine for Bodies-manual method/strip method.  Chemical Examination of urine for Bodies-manual method/strip method.  Chemical Examination of urine for ble pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examination of urine for bile pigments - manual method/strip method.  Chemical Examin	Area	Activity to be done.	Number of students
Accepting and rejecting criteria for samples.  Reception, Sample collection. Accepting and rejecting criteria for samples.  Main Lab Biochemistry Helping the senior technicians in estimation of Various analytes  1. Preparing the anticoagulants 7  Preparing the anticoagulants 7  Preparing the Leishman's stain.  3. Preparing peripheral blood smear.  4. Staining of peripheral blood smear.  5. Preparing and staining thick smear 6. Estimation of Haemoglobin by sahli's method and by photoelectric method.  7. Estimation of PCV by Micro and Macro Methods.  9. Physical examination of Urine.  10. Chemical Examination of urine for sugars-manual method/strip method.  11. Chemical Examination of urine for Potoens-manual method/strip method.  12. Chemical Examination of urine for Potoens-manual method/strip method.  13. Chemical Examination of urine for blood-manual method/strip method.  14. Chemical Examination of urine for blood-manual method/strip method.  15. Chemical Examination of urine for blood-manual method/strip method.  16. Preparing wet smear for urine for bile giments - manual method/strip method.  17. Chemical Examination of urine for bile giments - manual method/strip method.  18. Chemical Examination of urine for bile giments - manual method/strip method.  19. Preparing processing the blood sample for reticulocyte count by wet and dry method.  10. Charging the improved Neubauer chamber		.Accepting and rejecting criteria for samples.	
collection.Accepting and rejecting criteria for samples.  Main Lab Biochemistry Helping the senior technicians in estimation of Various analytes  2. Preparing the anticoagulants 2. Preparing the Leishman's stain. 3. Preparing peripheral blood smear. 4. Staining of peripheral blood smear. 5. Preparing and staining thick smear. 6. Estimation of haemoglobin by sahli's method and by photoelectric method. 7. Estimation of ESR 8. Estimation of PCV by Micro and Macro Methods. 9. Physical examination of Urine. 10. Chemical Examination of urine for sugars-manual method/strip method. 11. Chemical Examination of urine for Ketone bodiesmanual method/strip method. 12. Chemical Examination of urine for Protiens-manual method/strip method. 13. Chemical Examination of urine for Blood-manual method/strip method. 14. Chemical Examination of urine for blood-manual method/strip method. 15. Chemical Examination of urine for blood-manual method/strip method. 16. Preparing wet smear for urine for bile pigments - manual method/strip method. 17. Preparing wet smear for urine for bile pigments - manual method/strip method. 18. Charging the improved Neubauer chamber	Outpatient Pathology Lab	.Accepting and rejecting criteria for	
Main Lab Pathology	Outpatient Microbiology Lab	collection.Accepting and rejecting	7
2. Preparing the Leishman's statin. 3. Preparing peripheral blood smear. 4. Staining of peripheral blood smear. 5. Preparing and staining thick smear . 6. Estimation of haemoglobin by sahli's method and by photoelectric method. 7. Estimation of ESR 8. Estimation of PCV by Micro and Macro Methods. 9. Physical examination of Urine. 10. Chemical Examination of urine for sugars-manual method/strip method. 11. Chemical Examination of urine for Ketone bodiesmanual method/strip method. 12. Chemical Examination of urine for Protiens-manual method/strip method. 13. Chemical Examination of urine for Blood-manual method/strip method. 14. Chemical Examination of urine for blood-manual method/strip method. 15. Chemical Examination of urine for bile salts - manual method/strip method. 16. Preparing method. 17. Chemical Examination of urine for bile salts - manual method/strip method. 18. Chemical Examination of urine for bile pigments - manual method/strip method. 19. Preparing wet smear for urine microscopy. 19. Preparing processing the blood sample for reticulocyte count by wet and dry method. 18. Charging the improved Neubauer chamber	Main Lab Biochemistry	1 0	7
19. WBC Count by manual method. 20. Platelet Count by manual	Main Lab Pathology	<ol> <li>Preparing the anticoagulants</li> <li>Preparing the Leishman's stain.</li> <li>Preparing peripheral blood smear.</li> <li>Staining of peripheral blood smear.</li> <li>Preparing and staining thick smear.</li> <li>Estimation of haemoglobin by sahli's method and by photoelectric method.</li> <li>Estimation of ESR</li> <li>Estimation of PCV by Micro and Macro Methods.</li> <li>Physical examination of Urine.</li> <li>Chemical Examination of urine for sugars-manual method/strip method.</li> <li>Chemical Examination of urine for Ketone bodiesmanual method/strip method.</li> <li>Chemical Examination of urine for Protiens-manual method/strip method.</li> <li>Chemical Examination of urine for blood-manual method/strip method.</li> <li>Chemical Examination of urine for bile salts -manual method/strip method.</li> <li>Chemical Examination of urine for bile salts -manual method/strip method.</li> <li>Chemical Examination of urine for bile pigments -manual method/strip method.</li> <li>Chemical Examination of urine for bile pigments -manual method/strip method.</li> <li>Chemical Examination of urine for bile pigments -manual method/strip method.</li> <li>Chemical Examination of urine for bile pigments -manual method/strip method.</li> <li>Chemical Examination of urine for bile pigments -manual method/strip method.</li> <li>Chemical Examination of urine microscopy.</li> <li>Preparing wet smear for urine microscopy.</li> <li>Preparing the improved Neubauer chamber</li> <li>WBC Count by manual method.</li> </ol>	7

	method.  21. Processing of body fluids for cell count.  22. Processing of semen for sperm count.  23. Coagulation Tests- Bleeding time, Clotting time, Prothrombin time [PT]& APTT  24. Absolute Eosinophil count-Materials required, diluting fluids, procedure,  25. Sickle cell preparation.  26. Screening of blood donor.	
Main Lab Microbiology	<ul> <li>Reception area and its quality control.</li> <li>Cleaning and maintenance of equipment, glassware.</li> <li>Preparation of stains, culture media and sugar medias.</li> <li>Proper Disposal of lab waste.</li> </ul>	5

First month they will be taught about the theoretical aspects of the first 3 units of Biochemistry, Microbiology and Pathology. This can be within the premises of hospital during morning hours to orient them into appropriate subjects. Simultaneously practical aspects of Physiology recording BP, Temperature, Respiratory rate, TC, DLC can also be practiced.

OJT Master and lecture incharge should work in coordination with hospital superintendent.

Printed Log book should be designed and every day the student should be able to describe the activity done for that day. It should be signed by the concerned incharge staff in the respective wards/labs.

**Example**: If the student strength is 40 they can attend by rotation as below. For second year

Area	Activity to be done.	Number of students
Outpatient Biochemistry Lab	Reception,Sample collection.Accepting and rejecting criteria for samples.	2
Outpatient Pathology Lab	Reception ,Sample collection	2
Outpatient Microbiology Lab	Reception ,Sample collection	2
Inpatient G.Medicine	collection of samples, Recording of BP,Pulse ,Respiratory rate,	5
Inpatient G.Surgery	collection of samples, Recording of BP,Pulse ,Respiratory rate,	5
Inpatient Gynaecology	collection of samples, Recording of BP,Pulse ,Respiratory rate,	5
Inpatient Paediatrics	collection of samples, Recording of BP,Pulse ,Respiratory rate,	5
casualty	collection of samples, Recording of BP,Pulse ,Respiratory rate,	5
Main Lab Biochemistry	Helping the senior technicians in estimation of Various analytes	5
Main Lab Pathology	1. Performing Coombs Test – a) Direct b)	5

	indirect	
	2. Staining of Cytological smears.	
	3. Setting up of tray for bone marrow	
	aspiration	
	4. Preparing bone marrow smears –	
	methods- Imprints smears, crush smears.	
	5. Staining of bone marrow smear.	
	6. Buffy coat preparation.	
	7. Techniques of grouping and cross	
	matching	
	8. Receiving and preserving histopathology	
	specimens.	
	9. Maintaining the registers-receiving	
	register, grossing register, slide register,	
	report issue register.	
	10. Taking the consent for investigation.	
	11. Counseling of patient before and after	
	test.	
	12. Maintaining haematology analyser.	
	13. Processing the sample in haematology	
	analyser.	
	14. Fixation of Histopathology specimen.	
	15. Preparing the specimen for grossing.	
	16. Processing of histopathology specimen	
	manual and automated method.	
	17. Paraffin embedding and block making	
	18. Trimming of blocks	
	19. Tissue section cutting.	
	20. H&E Staining of tissue sections	
	21. H&E Staining cytology slides.	
	22. Mounting of slides	
	23. Maintaining microtome and tissue	
	processor and tissue floatation bath.	
	24. Sharpening of microtome knife.	
	25. Staining of histopathology sections.	
	26. Fixatives for cytology specimens.	
	27. Processing fluid sample for cytology.	
	28. Preparing tray for FNAC and guided	
	aspiration.	
	29. PAP stain preparation and staining of	
	smears.	
	30. Maintenance & Preservation of cytology	
	slides, histopathology blocks & slides.	
Main Lah Migrahiglagy	• Proposition of direct amount or 1	5
Main Lab Microbiology	Preparation of direct smears and striping of smears	5
	staining of smears.	
	Techniques of inoculation on media	
	and biochemical sugars for the	
	isolation of bacteria.	
	Stool concentration techniques and	
	microscopy of stool.	
	Serological tests.	
	<ul> <li>Antibiotic sensitivity tests.</li> </ul>	
	<ul> <li>Maintenance of stock cultures.</li> </ul>	
	Mycology processing	
		l l

VIII

## MODEL QUESTION PAPER MEDICAL LAB TECHNICIAN FIRST YEAR Paper – I: BIO-CHEMISTRY (THEORY)

Time: 3 Hours Max. Marks: 50

**SECTION-A** 

Note: (i) Answer all the Questions

(ii) Each Question carries 2 marks

10X2=20

- 1. Give the normal values of Blood sugars.
- 2. Define solution.
- 3. Mention the different methods of Blood collection.
- 4. What are the different types of Urine specimens? Give examples of urinary preservatives.
- 5. Explain the terms a) Solute b) solvent.
- 6. What are the hygroscopic substances? Give examples.
- 7. Expand GTT and give the normal values of serum uric acid.
- 8. Write the source of Vitamin-A. And write the diseases caused by its deficiency.
- 9. Write the names of water soluble vitamins.
- 10. Give the classification of carbohydrates.

#### **Section-B**

**Note: (i) Answer any five Questions** 

(ii) Each Question carries 6 marks

5X6=30

- 11. Give the classification of Lipids and write the biological importance.
- 12. Write the determination of Blood glucose using GOD-POD method.
- 13. Give a note on different types of Glass ware used in Bio-chemistry lab. Write the applications.
- 14. Describe the prevention, safety and first- Aid in lab accidents.
- 15. Write the principle and construction of Electrical Centrifuge. And give the applications.
- 16. Define Colorimetry. Describe the construction, operation and uses of colorimeter.
- 17. What is Phlebotomy? Describe the collection of venous blood.
- 18. Write the principle, construction and applications of Spectro photometer.

#### MEDICAL LAB TECHNICIAN

#### **FIRST YEAR**

#### Paper – II: MICRO-BIOLOGY & PATHOLOGY (THEORY)

Time: 3 Hours Max. Marks: 50

## **SECTION-A**

Note: (i) Answer all the Questions

## (ii) Each Question carries 2 marks

10X2=20

- 1. Mention the names of different body fluids.
- 2. Write the physical properties of Urine.
- 3. Write the names of Bile salts and Bile Pigments.
- 4. Give the normal values of Total RBC & Platelets.
- 5. Write about a) Glycosurea b) Hematuria.
- 6. What is Liquefaction time of Semen?
- 7. Define Sterilization and disinfection.
- 8. Give the differences between Gram Positive & Gram Negative bacteria
- 9. Write the principle of Compound Micro-scope
- 10. Write the contributions of Antony van Leeuwenhoek&Louis Pasteur to Microbiology

### **SECTION-B**

## **Note: (i) Answer any five Questions**

## (ii) Each Question carries 6 marks

5X6 = 30

- 11. What is an Anti-Coagulant? Write the uses , Quantities required and preparation of EDTA & Sodium citrate
- 12. Describe the Estimation of ESR. Give the normal values of and write the clinical importance.
- 13. Write the Qualitative determination of Urine sugar. And write the clinical importance.
- 14. What are the collection methods of Semen? Describe the Physical examination of Semen.
- 15. Describe the construction & operation of Fluorescent Microscope and write the applications.
- 16. What are the different methods of Sterilization? Describe the construction and operation of Autoclave
- 17. Write a note on Media for Blood cultures and anaerobic media. Write the composition & preparation of Zeil Nelsons stain.
- 18. Estimation of Hemoglobin by Sahli's method.

## MEDICAL LAB TECHNICIAN FIRST YEAR

## Paper – III: ANATOMY & PHYSIOLOGY (THEORY)

Time: 3 Hours Max. Marks: 50

## **SECTION-A**

Note: (i) Answer all the Questions

(ii) Each Question carries 2 marks

10X2=20

- 1. Define Anatomy & physiology
- 2. Mention the varieties of tissues in our body
- 3. Write the functions of saliva
- 4. What is Dextro Cardia
- 5. List the Proximal Row Carpal Bones
- **6.** What is Meningitis
- 7. What are the Exocrine Glands
- **8.** Write the surfaces of Kidney
- **9.** Mention the parts of Fallopian Tubes
- 10. Write names of the following Nerves & arteries
  - a) 7<sup>th</sup> Cranial nerve
- c) 4<sup>th</sup> Cranial nerve
- b) Blood supply to liver and Heart

### **SECTION-B**

Note: (i) Answer any five Questions

(ii) Each Question carries 6 marks

5X6=30

- 11. Draw the Neat and labeled diagram of heart and explain coronary circulation
- 12. Write the classification of Bones with examples and mention the functions of Bones
- 13. Define organ write the organs present in Respiratory system and explain one in detail
- 14. Draw the diagram of skin and write the functions of skin
- 15. Draw the labeled structure of stomach and explain the functions of liver
- 16. Write the composition of blood and functions of blood
- 17. a) list the Endocrine glands
  - b) Write short notes on
    - 1) Uterus 2) Tonsils
- 3) Appendix
- **18.** Draw a neat diagram of urinary system with labelling. And explain the formation of urine.

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR

### **Paper – I: BIO-CHEMISTRY (THEORY)**

Time: 3 Hours Max. Marks: 50

## **SECTION-A**

**Note: (i) Answer all the Questions** 

(ii) Each Question carries 2 marks

10X2=20

- 1. What are transaminases? Give examples.
- 2. Write the clinical application of chromatography and electrophoresis
- 3. Write the principle of Flame photometry
- 4. Define a) Glycolysis b) Urea cycle
- 5. What is quality assurance? Explain internal quality control
- 6. Mention Lipid profile tests
- 7. What are the Thyroid Hormone assayed by RIA
- 8. Give the Normal values of Serum Bilirubin & Total Proteins
- 9. Define enzymes. Mention the unit of measurement.
- 10. Mention various tests done to asses renal functioning

## **SECTION-B**

### **Note: (i) Answer any five Questions**

(ii) Each Question carries 6 marks

5X6=30

- 11. Write about determination of Glycosylated hemoglobin and its clinical importance
- 12. Write about enzyme. Give the classification
- 13. Determination of Serum total Bilirubin
- 14. Describe the construction, operation, application of a Flamephotometer with a neat schematic diagram
- 15. How do you determine serum amylase? Give the principle requirements and methods
- 16. Discuss about automation in a biochemical laboratory
- 17. Define primary standard and secondary standard classify different titrimetric methods
- 18. Explain separation of plant pigments by paper chromatography

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR

## Paper – II: MICRO-BIOLOGY (THEORY)

Time: 3 Hours Max. Marks: 50

**SECTION-A** 

Note: (i) Answer all the Questions

## (ii) Each Question carries 2 marks

10X2=20

- 1. Expand ELISA & RIA.
- 2. Name causative Organism for Cholera and Diphtheria.
- 3. Define Antigen and Antibody.
- 4. Give the importance of Vaccines.
- 5. Explain about Euthanasia.
- 6. Write difference between Gram positive & Gram negative bacteria.
- 7. Define Immunity
- 8. Principle of stock culture
- 9. Morphology of Gonococci
- 10. Importance of Mycology

## **SECTION-B**

Note: (i) Answer any five Questions

#### (ii) Each Question carries 6 marks

5X6 = 30

- 11. Write Morphology and lab diagnosis of E. histolytica
- 12. Antibiotic sensitivity Test
- 13. Explain the Normal flora of Micro-organism in Human body
- 14. Write about Bacteria causing food poisoning.
- 15. Write Short Notes on
  - 1) Candida
  - 2) Penicillin
  - 3) Actinomyces.
- 16. Write a note on Preservation, methods of stock culture and their importance
- 17. Write about the collection and processing of faecal samples.
- 18. Write morphology, cultural characteristics and lab diagnosis of E.coli

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR

## **Paper – III: PATHOLOGY (THEORY)**

Time: 3 Hours Max. Marks: 50

**SECTION-A** 

Note: (i) Answer all the Questions

### (ii) Each Question carries 2 marks

10X2=20

- 1. Mention specimen collection sites for Bone Marrow Examination.
- 2. Expand PAS and APTT.
- 3. Write the principle for sickle cell preparation.
- 4. Expand CT & BT.
- 5. Define Biopsy.
- 6. Write the names of various Microtomes.
- 7. Explain the characters of Blood donor.
- 8. What is Regressive stain?
- 9. What is Mordant?
- 10. Write the composition of Leishman stain.

#### **SECTION-B**

**Note: (i) Answer any five Questions** 

### (ii) Each Question carries 6 marks

5X6=30

- 11. Explain automatic tissue processing.
- 12. Give blood grouping procedure and its importance.
- 13. Identification of Microfilaria.
- 14. Write the principle and clinical signification of sickle cell preparation
- 15. Describe the Decalcification by nitric acid method
- 16. Explain about the Direct & indirect comb's test. Give clinical importance
- 17. Write the principle, procedure & material required for L.E. Cell test
- 18. Give a note on preparation of Thick & Thin blood films and uses

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER FIRST YEAR PAPER – I: BIO-CHEMISTRY (PRACTICAL)

Time: 3 Hours Max. Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks 2X15=30

- 1. Collect the Venous blood. Write the procedure. Show the result
- 2. Prepare 250ml of 0.9% Nacl solution. Show the preparation. write the procedure
- 3. Find out the Glucose in the given urine sample by Benedict's method. Write the procedure. produce the results

## **SECTION-B**

4. Spotters 10marks

## SECTION -C

5. Viva6. Record5marks5marks

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER FIRST YEAR

## Paper – II: MICRO-BIOLOGY & PATHOLOGY (PRACTICAL)

Time: 3 Hours Max.Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks 2X15=30

- 1. Estimate the Hb% in the given blood sample by Sahli's method. Produce the result. Write the procedure
- 2. Count the Total WBC in the given sample. Show the result. Write the procedure
- 3. Prepare Mac Conkey's agar media. Show the preparation. Write the procedure.

## **SECTION-B**

4. Spotters 10marks

## SECTION -C

5. Viva6. Record5marks5marks

# MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER FIRST YEAR Paper –II: ANATOMY & PHYSIOLOGY (PRACTICAL)

Time: 3 Hours Max.Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks

2X15=30

- 1. Count the Total RBC in the given blood sample using Neubauer counting chamber. Show the result. Write the procedure
- 2. Draw the neat diagrams of Skull & Humerus with labeling
- 3. Estimate the Blood pressure. Write the procedure

## **SECTION-B**

4 .Spotters 10marks

## **SECTION -C**

5. Viva 5marks
6. Record 5marks

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR

PAPER – I: BIO-CHEMISTRY (PRACTICAL)

Time: 3 Hours Max.Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks 2X15=30

- 1. Estimate the serum sodium in the given sample using FPM. Produce the result. write the procedure
- 2. Determine the serum Bilirubin in the given sample. Show the result. Write the procedure.
- 3. Estimate the concentration of blood urea in the given sample by DAM-TSC method. Show the result. Write the procedure

## **SECTION-B**

4 .Spotters 10marks

## **SECTION -C**

5.Viva 5marks
6. Record 5marks

# MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR PAPER –II: MICRO-BIOLOGY (PRACTICAL)

Time: 3 Hours Max.Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks 2X15=30

- 1. Perform widal test with the given sample. Show the result and write the procedure.
- 2. To identify the Ova & Cyst in the given sample, perform concentration technique. Show the result and write the procedure
- 3. Put up the Biochemical reactions required to identify Escherichia coli. Write the procedure.

## **SECTION-B**

4 .Spotters 10marks

## SECTION -C

5. Viva 5 marks 6. Record 5 marks

## MEDICAL LAB TECHNOLOGY MODEL QUESTION PAPER SECOND YEAR Paper –III: PATHOLOGY (PRACTICAL)

Time: 3 Hours Max.Marks: 50

**SECTION -A** 

Note: Attempt any two practicals. Each question carries 15marks 2X15=30

- 1. Find out the Blood Group in the given sample. Show the result and write the procedure
- 2. Find out the Clotting time & bleeding time in your own blood. Show the result and write the procedure
- 3. Describe the Sharpening of Microtome Knives.

## **SECTION-B**

4 .Spotters 10 marks

## SECTION -C

5. Viva 5 marks 6. Record 5 marks

## IX LIST OF EQUIPMENTS

## BIOCHEMISTRY

1.	Hot Plate	1
2.	Gas Cylinder with Burner	2
3.	Spirit Lamps	15
4.	Electrical Centrifuges	2
5.	Refrigerator 165 lit.	1
6	Colorimeter	1
7.	Hot Air oven	1
8.	Water bath	1
9.	Simple balance	1
10.	Electronic balance	1
11.	Flame photometer	1
12.	Spectrophotometer	1
13.	Flourimeter	1
14.	Incubator	1
15.	Electrophoresis apparatus	1
16.	Computer with Printer	1
17.	Semi Auto Analyzer.	1
18.	Automated Electrolyte Analyzer	1

## **GLASSWARE**

1.	Test Tubes	
1.	18X150mm	100
	15X150mm	100
	15X125mm	100
2.	Centrifuge Tubes	20
3.	Beakers- 250 ml	5
4.	Pipettes	
т.	a)Volumetric Pipettes.	
	2ml	5 No's
	5ml	5 No's
	10ml	5 No's
	20ml	5 No's
	25ml	5 No's
	b)Serological Pipettes.	31103
	1ml 1/100	5 No's
	2ml/1/100	5 No's
	5m1/100	5 No's
	1.10 ml1/10	5 No's
	2ml 1/10 ml	5 No's
	0.1ml 1/100ml	5 No's
	02 ml 1/100	5 No's
	C)Ostwald pipettes	
	0.1ml	2
	0.2ml	2
	0.5 ml.	2
5	Burettes	
	25 ml	5
	50 ml	5
6	Reagent Bottles	
	60 ml	10
	120 ml	10
	250 ml	10
	500 ml	10
	1000 ml	10
7.	Dropper Bottles 30 ml	5

8.	Watch glass	6
9.	Volumetric Flasks	
	25ml	5
	50ml	5
	100ml	5
	250ml	5
	500ml	5
	1000ml	3
10.	Stoppaerd graduated Test Tubes	
	15ml	5
	40ml	5
	50ml	5
11.	Distillation assembly [complete set]	1
12.	Round Bottom flask 500 ml & 1000 ml	1+1
13.	Filter paper	1 Ream
14.	What man filter paper	No.1 - 20
		sheets
	What man filter paper	No.2- 10
		sheets.
15.	Cotton ( absorbent)	1 kg.
16.	Glass slides	100
17.	Plastic wash bottles 500 ml	10
18.	Mortar and Pestle	2 no's
19	Measuring Jar	
20	Test tube racks, Test tube holders and Funnels	

## MICROBIOLOGY

1.	Compound Microscope	
2.	Variable Volume Micro Pipettes Full Set	1
3.	Centrifuge	1
4.	Refrigerator	
5.	Autoclave	
6.	Hot air oven	
7.	Incubator Bacteriological	
	BOD Incubator	
	Micro Plate Washer and Reader	
8.	Distil water plant	
9.	Pipette washer	
10.	Anaerobic Jar	
11.	Vacuum Pump	
12.	Analytical Balance	
13.	Water Bath	
14	VDRL Rotator	
15	Bunsen Burner with Gas Connection	
16	Inoculating loop	
17	Straight wire	
18	Petri dishes 100X17	100 no's
19	Filter paper	1 Ream
20	What man filter paper	No.1 – 20
		sheets
21	Dropper Bottles 30 ml	5
22	Cotton ( absorbent)	1 kg.
23.	Glass slides	100
24	Depression Slides	
25	Cover Slips	
26	Serum Storage Vials and Boxes	
27	Test Tubes 150X19	100

28	100X12	100
29	Pipettes	
30	10ml	10 Nos.
31	5ml	10 Nos.
32.	1ml	10 Nos.
33.	Wash bottles	5
34.	Spatulas	12
35	Reagent bottles	10
36	Measuring Cylinders 50 ml	5
37	Compound Microscope	5

## **PATHOLOGY**

1	D' 1 M'	1
1.	Binocular Microscope	1
2.	Hot air oven	1
3.	Incubator	1
4.	Centrifuge	1
5.	Haematology analyser (Blood cell counter ) 3/5 Part	1
6.	Water Bath	1
7.	Chemical balance	1
8.	Hot plate	1
9.	Stopwatch	1
10.	Haemometer	5
11.	Haemocytometer	5
12.	Lab Counter for DC	4
13.	ESR Stand	5
14.	ESR Tubes	5
15.	Motor and pestle	5
16	Urino meter	2
17	Microhaematocrit centrifuge	1
18	Automated pippets	5
19.	WBC pippets RBC pippetsHbpippets	20 each
20.	Spirit lamp	2
21	Syringes	
	20ml	10
	10ml	10
	5ml	10
	2ml	10
22	Beaker	
	100ml	5
	250ml	5
23	Test Tubes	
	10 ml	50
	15 ml	50
24	Watch glasses.	5
25	Trays	5
26.	Syringe dispenser to crush needles.	1
27.	Refrigerator	1
28.	Glass slides and cover slips.	50
29.	Tissue processor	1
30.	Microtome	1
31.	Microtome knives	3
32	Coupling jars	20
33.	Slide holder baskets	4
34	Water bath	1

## **MODEL LOG BOOK FOR MLT STUDENTS**

Date	Description of the work performed	Signature of the OJT Master /Lecturer	Classroom Lectures
6.5.17	Went to Hospital Biochemistry lab; Number of patients interacted ;		
8.5.17	Blood drawn from Number of patients		
9.5.17	Assisted in labelling the samples.		
10.5.17	Assisted in entering of samples in the register.		

Date	Description of the work performed	Signature of the OJT Master /Lecturer	Classroom Lectures
6.5.17	Went to Hospital microbiologylab Number of patients interacted ;		
8.5.17	Number of samples accepted, rejected entered		
9.5.17	Assisted in labelling the samples.		

## A. Collaborating Institutions for Curriculum transaction

- 1. All Hospitals.
- 2. All Medical Colleges.
- 3. All the National Laboratories
- 4. Regional Research Laboratories.
- 5. University Departments.
- 6. Pharmaceutical Companies and Educational Institutes.

## **B.** On the Job Training Centers.

- 1. Government Hospitals
- 2. PHCs Primary Health Centers.
- 3. Dispensaries.
- 4. Medical Colleges.
- 5. Private Hospitals
- 6. Private Labs.

## X. Qualification of Lecturers

- 1.MD Pathology / Microbiology / Biochemistry / DCP.
- 2. M. Sc Microbiology / Biochemistry / MLT.
- 3. B. Pharmacy/ MSc Genetics.
- 4. MBBS/MBBS(Hom)/BHMS
- 5. PGD. Clinical Bio chemistry.

## XI. Vertical Mobility

## A) With Bridge Course

- 1. B. Sc (BZC)
- 2. Courses through EAMCET

## B) Without Bridge Course.

- 1. B. Sc MLT, B.A & B.Com, D.MLT
- 2. B. Sc Microbiology
- 3. B. Sc Biochemistry
- 4. B. Sc Biotechnology
- 5. M.Sc. MLT/Biochemistry /Microbiology/ Biotechnology [at P.G. level]

## XII REFERENCE BOOKS

#### **Biochemistry**

1. Harold Varley

#### **Anatomy & Physiology**

- 1. C.C. Chatterjee
- 2. Chowrasia

#### Microbiology

- 1. A manual on Medical laboratory Technicians.. A.v. Naigonkar
- 2. RamnikSood

## **Pathology**

- 1. RamnikSood
- 2. Mukerjee 3 volumes
- 3. Talib
- 4. PrafulGodkar
- 5. WHO Lab Manual
- 6. Harsh Mohan Practical book.
- 7. Tejinder Singh Practical book.
- 8. Praful Godkar
- 9. K.M. Samuel.

## XIII. LIST OF SUBJECT COMMITTEE MEMBERS:

	D 11 D 1 1/D
1.	Dr. Ather Fatima, M.D.,
	Associate Professor,
	Department of Pathology,
	Govt Medial College,
	Nizamabad.
2.	Dr. M. Rama Devi, M.D.
	Professor and HOD,
	Department of Biochemistry
	Osmania Medical College,
	Hyderabad.
3.	Dr. Pooja Palla,
3.	Assistant Professor
	Department of Microbiology.
	Govt. Medial College,
	Nizamabad
4.	Sri. S. Srinivasa Rao
	Principal
	GJC, Pidiprolu
	Khammam
Verifie	d & Corrected By
1.	Dr. Shravan Kumar
	Prof &Hod of Pathology
	Gandhi Medical College,
	Hyderabad
2.	Dr. Nagamani
	Prof &Hod of Microbiology
	Gandhi Medical College,
2	Hyderabad
3.	Dr. Suleman
	Prof of Bio Chemistry
	Gandhi Medical College, Hyderabad
Co. ord	linator:-Sri K.Vishweshwar,
Co- ord	O/o the Commissioner of Intermediate Education,
Andhra Pradesh,	
	Hyderabad
ĺ	11, 4014044

Sd/- Dr A. Ashok COMMISSIONER OF INTERMEDIATE EDUCATION